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Book • 2021Advanced Methods in Molecular Biology and Biotechnology: A Practical Lab Manual is a concise reference on common protocols and techniques for advanced molecular biology and biotech ... read full descriptionSelect all / Deselect all Explores a wide range of advanced methods that can be applied by researchers in molecular biology and biotechnology Features clear, step-by-step instruction for applying the techniques covered Offers an introduction to laboratory protocols and recommendations for best practice when conducting experimental work, including standard operating procedures for key equipment Explores a wide range of advanced methods that can be applied by researchers in molecular biology and biotechnology Features clear, step-by-step instruction for applying the techniques covered Offers an introduction to laboratory protocols and recommendations for best practice when conducting experimental work, including standard operating procedures for key equipment You currently don't have access to this book, however you can purchase separate chapters directly from the table of contents or buy the full version.Purchase the book BiologyTypeOptional (OP)Language of instructionHave oral and written skills in English for communicating results, conclusions and processes deriving from research in the field of biology Manipulate databases used in the field of biology. Master basic laboratory techniques in biology, apply protocols and use appropriate instruments, observing safety norms and correctly interpreting the results obtained.Understand the processes of functional integration in organisms based on knowledge of subcellular structure and organism cell types. Students can apply their knowledge to their work or vocation in a professional manner and have competencies typically demonstrated through drafting and defending arguments and solving problems in their field of study. Students have demonstrated knowledge and understanding in a field of study that builds on general secondary education with the support of advanced textbooks and knowledge of the latest advances in this field of study. Be a critical thinker before knowledge in all its dimensions. Show intellectual, cultural and scientific curiosity and a commitment to professional rigour and quality. Use oral, written and audiovisual forms of communication, in one's own language and in foreign languages, with a high standard of use, form and content. The Molecular Biology Techniques subject is taught in a project-based learning (PBL) format. The experimental project consists in the identification of an organism through the DNA barcoding technique, which is based on the amplification and sequencing of a mitochondrial gene: Cytochrom oxidase I (CO1). Throughout the project, several molecular biology techniques are introduced, discussed and applied, while reinforcing different technical and cross-curricular skills. Subject aims Observe at all times the safety and operation regulations in the laboratory. Manipulate properly the routine-use laboratory instruments and correctly apply microbiology and molecular biology protocols. Record in a proper and orderly manner all the activities carried out in the laboratory. Understand and successfully apply protocols for extraction, amplification and analysis of nucleic acids. Carry out the molecular identification of a species of insect from a tissue sample. Sustainable Development Goals. This subject addresses the Study of Global and Local Biodiversity in the context of the following SDGs: Goal 14: Life Below Water - Conserve and sustainably use the oceans, seas and marine resources Goal 15: Life On Land - Biodiversity, Forests, DesertificationLearning outcomesLearning outcomes Gains autonomy and initiative in the laboratory. Properly uses routine-use instruments in a biological laboratory, including safety and disposal regulations. Plans the execution and carries out an experimental protocol in a team-work context and in a suitable time. Keeps an adequate activity record and issue reports that justify and analyze the work done. Performs a critical interpretation of the experimental results to draw reasonable conclusions. Understands the basics of basic molecular biology techniques and applies them correctly. Searches the necessary bibliographical resources. Understands and is able to communicate complex oral and written messages. Writes written reports and documents with proper spelling and grammar. Content Introduction to laboratory work: Basic laboratory regulations. Reagent manipulation and waste disposal. Team work organization Introduction to the Barcode of Life Project: Molecular-based identification of species. Sample collection. Gathering and organization of sample metadata. Amplification and sequencing of cytochrome oxidase I (CO1): PCR amplification. Sanger sequencing. Data analysis: Data pre-processing and quality check. Sequence alignment and identification. Introduction to the phylogenetic analysis of sequences. EvaluationAssessment activities (which represent 100% of the final mark (FM)) Activity 1: Written test (40% of the FM). Minimum grade: 4/10. Resit allowed Activity 2: Team work public exposition (15% of the FM). Resit not allowed. Activity 3: Exercises and final report (30% of FM). Resit not allowed (late submission penalizes 20%). Activity 4: Personal performance and attitude in the laboratory (15% of the FM). Resit not allowed. Additional considerations Attendance at all sessions is mandatory. Justified absence up to a maximum of 20% of sessions. Unjustified absence to more than 20% of the sessions (or justified absence to more than 40%) results in a grade of 0/10 of activity 4. Absence to more than 40% of the practical activities implies failing the subject. The lack of punctuality will be assessed negatively and, if it is repeated and unjustified, it will be considered absence. Activity 2: Unjustified absence to final exposition results in a grade of 0/10 and a penalty of 25% on the grade obtained by the group to which the student belongs. Activity 4 will assess the following aspects: Proper work in the laboratory and correct use and care of basic materials and techniques. Understanding and correct application of the laboratory protocols. Results obtained in the practical experiments. Methodology The methodology of the project is based on the development of an eminently hands-on laboratory activity. Various resources and activities, both guided and self-study, are contemplated, which must allow students the adequate achievement of the competencies, skills and knowledge associated with the protocols that will be developed throughout the sessions. BibliographyBasicW. John Kress and David L. Erickson, Editors (2012). DNA Barcodes: Methods and Protocols (1 ed.). Springer.Further readingTeachers will provide complementary bibliography and compulsory reading throughout the course via the Virtual Campus. An introduction to data integration and statistical methods used in contemporary Systems Biology, Bioinformatics and Systems Pharmacology research. The course covers methods to process raw data from genome-wide mRNA expression studies (microarrays and RNA-seq) including data normalization, differential expression, clustering, enrichment analysis and network construction. The course contains practical tutorials for using tools and setting up pipelines, but it also covers the mathematics behind the methods applied within the tools. The course is mostly appropriate for beginning graduate students and advanced undergraduates majoring in fields such as biology, math, physics, chemistry, computer science, biomedical and electrical engineering. The course should be useful for researchers who encounter large datasets in their own research. The course presents software tools developed by the Ma'ayan Laboratory ( from the Icahn School of Medicine at Mount Sinai, but also other freely available data analysis and visualization tools. The ultimate aim of the course is to enable participants to utilize the methods presented in this course for analyzing their own data for their own projects. For those participants that do not work in the field, the course introduces the current research challenges faced in the field of computational systems biology. The purpose of this chapter is to outline some of the common recombinant DNA methods in use today. These techniques are usually employed to isolate a defined portion of the genome, mostly a gene, from an organism or tissue of interest and, thereafter, to characterize the structure and function of this genetic material. To isolate a gene, genomic DNA is extracted from a selected tissue. For a better handling the relatively large DNA molecules are cut into a mixture of fragments by restriction endonucleases. The fragments are then separated from each other according to their size by gel electrophoresis. A procedure called Southern blotting is used to verify the presence of the desired gene in one of the DNA fragments separated on an agarose gel. The DNA fragments are transferred from the gel to a filter whereby the original fragment pattern is maintained. Then, a single-stranded DNA or RNA probe specific for the gene to be isolated is hybridized to its target fragments fixed to the filter. A radioactive or fluorescent tag is attached to the probe for subsequent identification. In cases where only transcribed sequences are to be isolated cytoplasmic messenger RNA (mRNA) is prepared instead of DNA. Analysis of RNA by a technique similar to Southern blotting is termed Northern blotting. Preservation of DNA sequences is usually achieved by DNA cloning. DNA cloning involves the insertion of a DNA fragment into a DNA vector and the stable incorporation of the recombinant DNA into a suitable host. Propagation of the host facilitates the amplification of the recombinant DNA for subsequent analysis.(ABSTRACT TRUNCATED AT 250 WORDS)

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